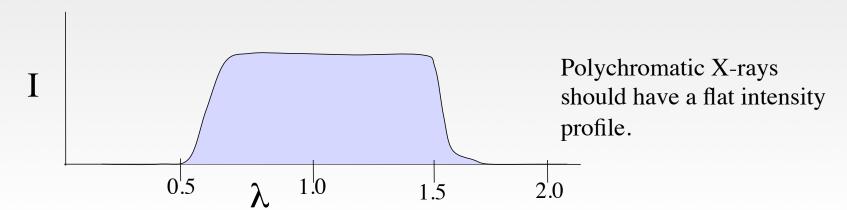
PSD '18 Xray lecture 12

cutting-edge crystallography

Laue photography,
X-ray laser,
Microcrystal Electron Diffraction

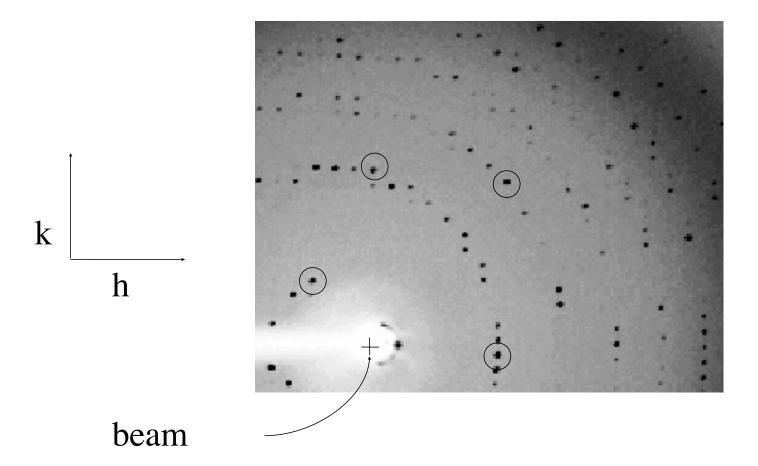
Laue Method

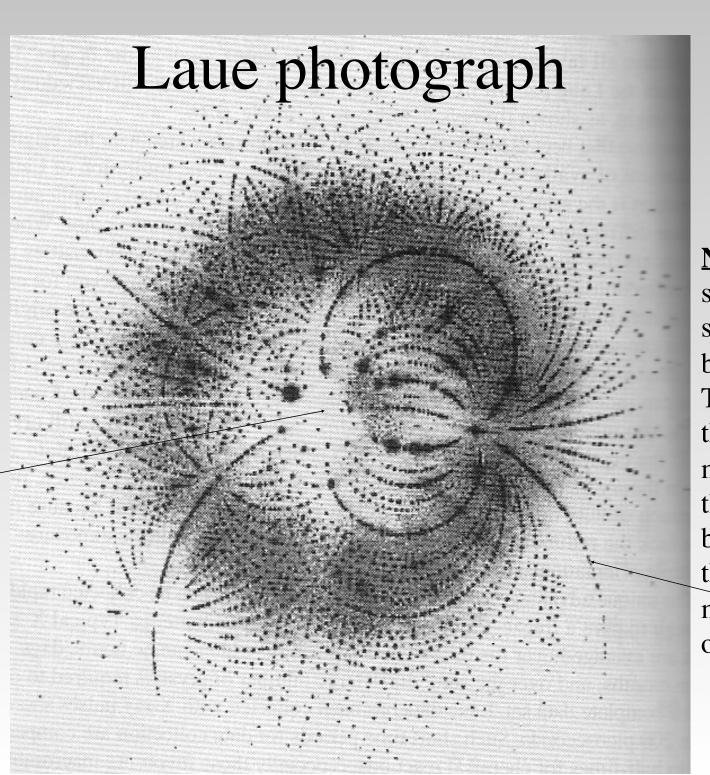
- •Uses polychromatic ("white") X-rays. (generally $\lambda < 2.0 \text{Å}$)
- •Allows data to be collected ultra-fast, leading to its application in "time-resolved" crystallography.
- •Still photographs cover a wide range of reciprocal space. A whole dataset may be collected on a few films.
- •Requires synchrotron radiation



Monochromatic diffraction

still image

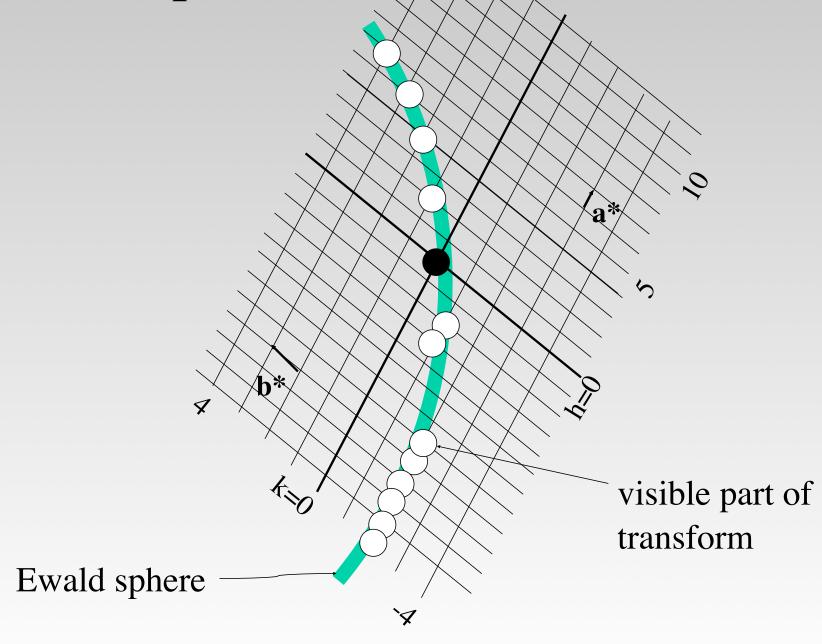


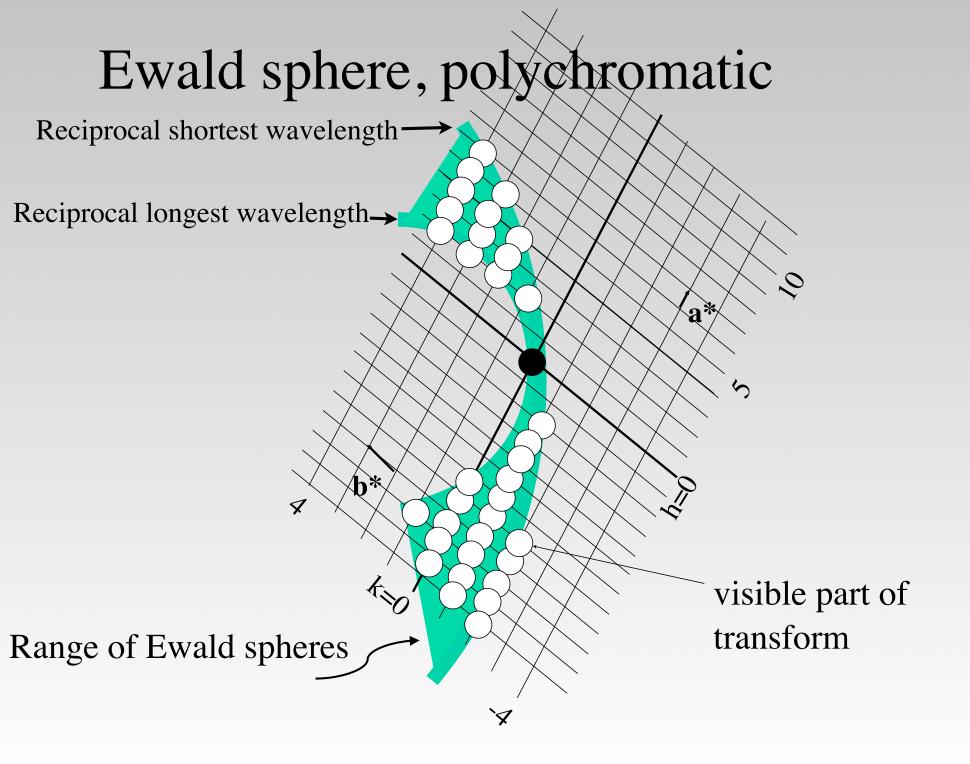


beam

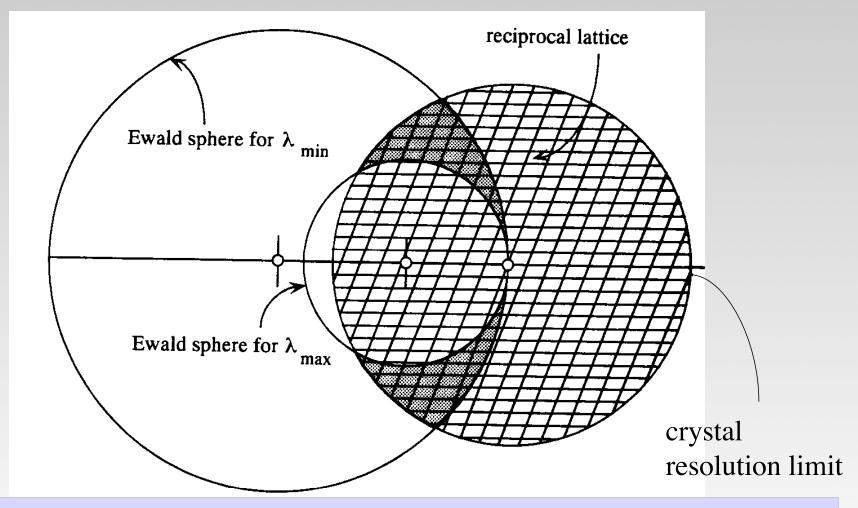
Note: Very small separation between spots. This means the crystals must be small, the beam must be small, and the crystals must be wellordered.

Ewald sphere, monochromatic



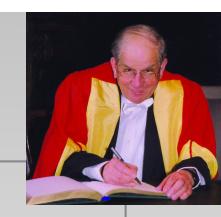


The Ewald sphere(s), polychromatic



Ewald sphere has radius $1/\lambda$. Longer wavelength X-rays cause the Ewald sphere to shrink, picking up a different part of the reciprocal lattice.

Cruikshank's dilemma



In monochromatic crystallography: one θ angle = one d. In Laue crystallography, one θ angle = a range of d's.

Bragg's Law for monochromatic

$$n\lambda = 2d \sin \theta$$

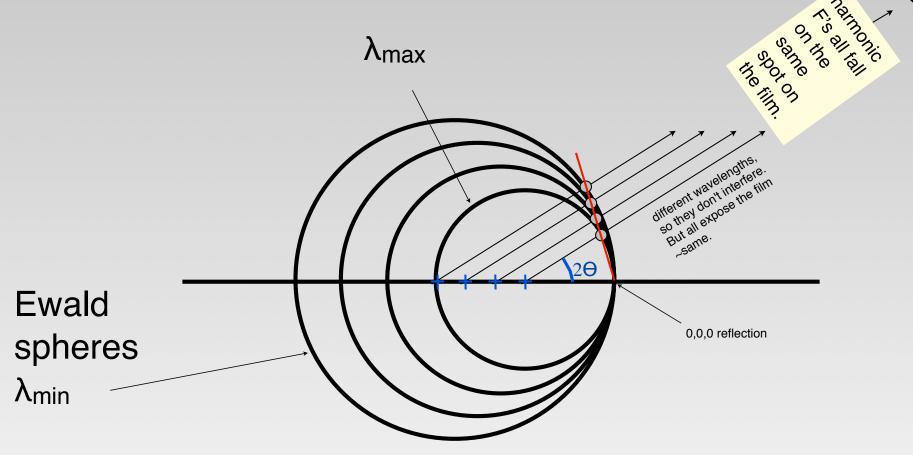
Bragg's Law(s) for Laue

$$n\lambda_{max} = 2d \sin \theta$$

$$= n\lambda_{max} / 2 = 2(d/2) \sin \theta$$

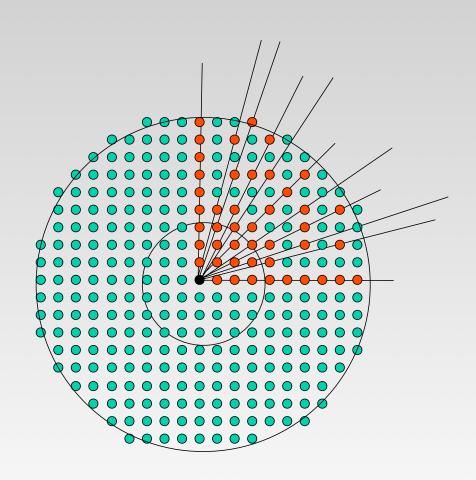
$$= n\lambda_{max} / 3 = 2(d/3) \sin \theta$$

harmonic reflections



Harmonic reflections (nh,nk,nl) have the same S direction, but the length is inversely proportional to λ . So (h,k,l) at λ =2.0Å and (2h,2k,2l) at λ =1.0Å diffract to exactly the same spot on the film.

Harmonics are reflections that lie along rays going out from the reciprocol space origin

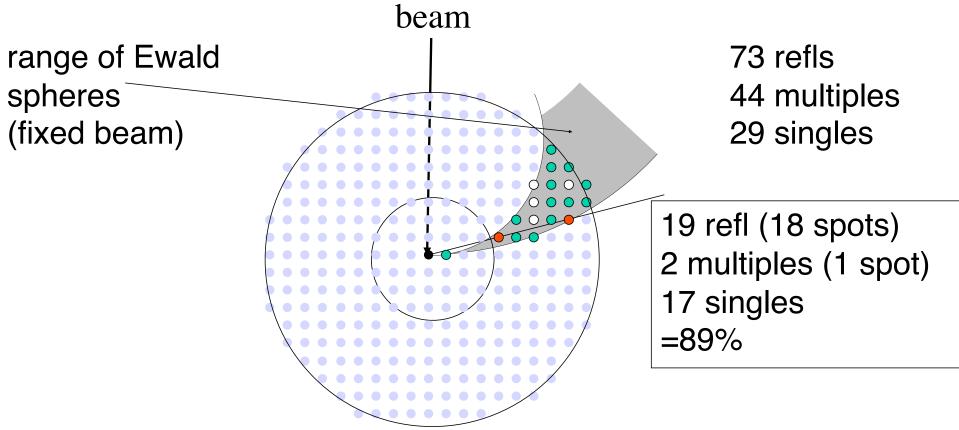


73 refls

44 multiples

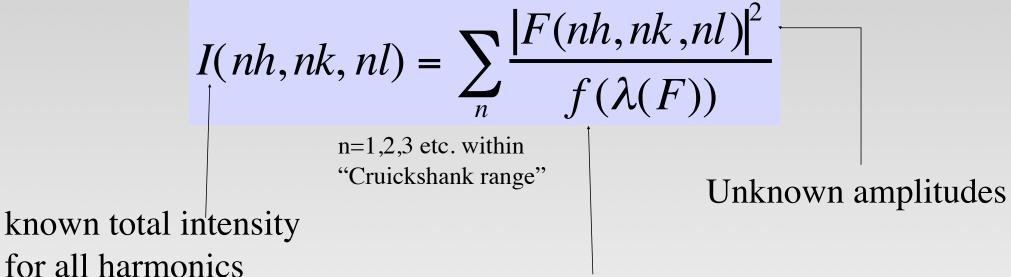
29 singles

Not all harmonics fall within the Ewald zone (Cruikshank range)



Now cut down all the trees except the ones that are on the Ewald sphere for one of the wavelengths (range λ_{min} - λ_{max}). Only the trees in the grey region remain. How many are multiples?

Combining harmonics



Scale factor for each.

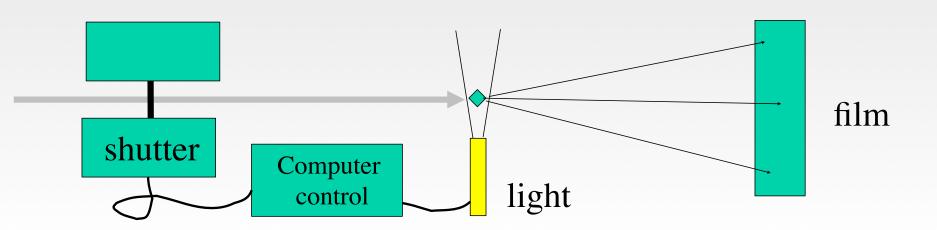
Accounts for beam intensity, polarization.

Time-resolved crystallography!

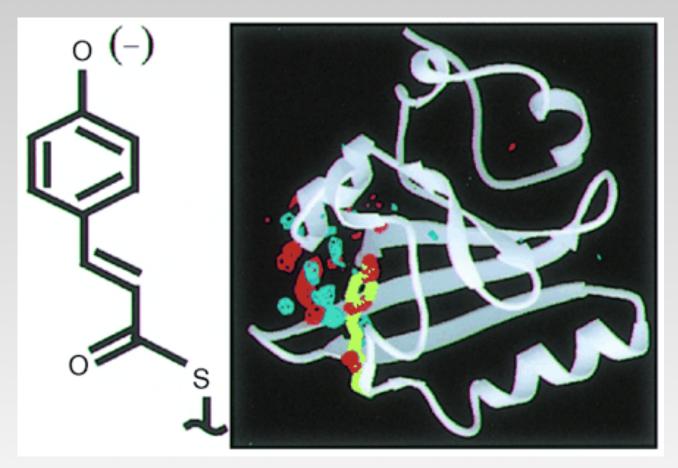
If a reaction can be initiated in a crystal, simultaneously throughout the crystal, then Laue photography can capture the structural changes at the ns (10-9 s) to ms (10-3 s) timescale.

Crystals must withstand ultrahigh fluxes of X-rays, or are destroyed in the process.

Light-initiated reactions can be studied using the Laue method.



Photoactive yellow protein



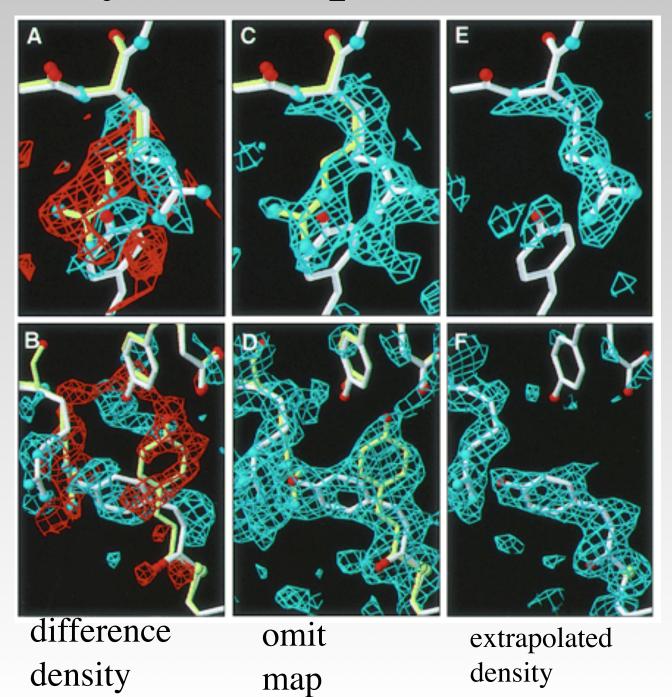
4-hydroxycinnamyl chromophore

Photoactive yellow protein (from the phototrophic bacterium Ectothiorhodospira halophila)

Genick et al. Science, 275 (5305): 1471

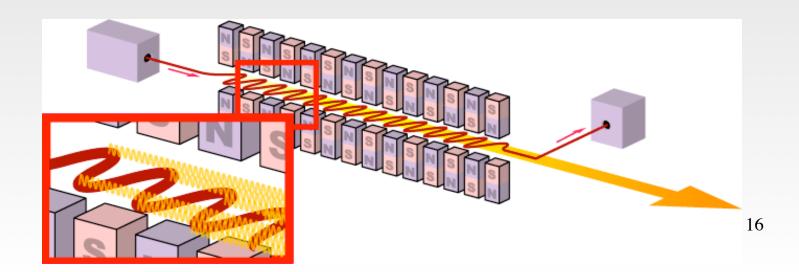
Hybrid maps

F_{bleached} - F_{dark}



X-ray free electron laser (XFEL)

- First successful X-ray laser, published in July 2012.
- Generated by an electron beam which moves freely through a magnetic structure, under vacuum.
- Tunable to a wide range of frequencies.
- Billions of times more intense than conventional methods.

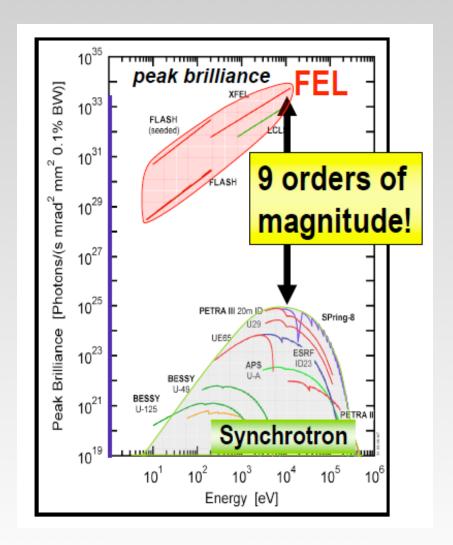


European FEL construction site, Hamburg

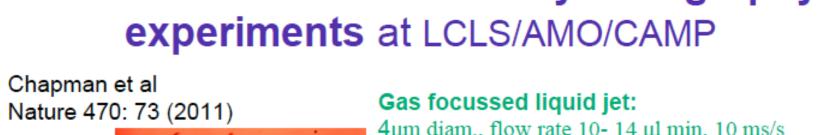


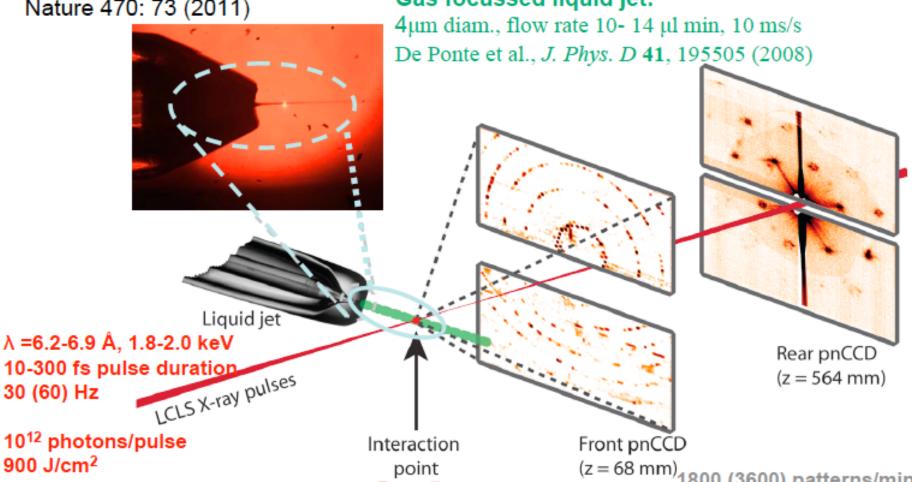
Free electron laser is quantum leap in brilliance

- 10¹²⁻¹³ photons: ~ 10 fs pulses
- repetition rate: now 120 Hz
- photon energies: 10 keV
- transversally: fully coherent



First serial femtosecond crystallography experiments at LCLS/AMO/CAMP





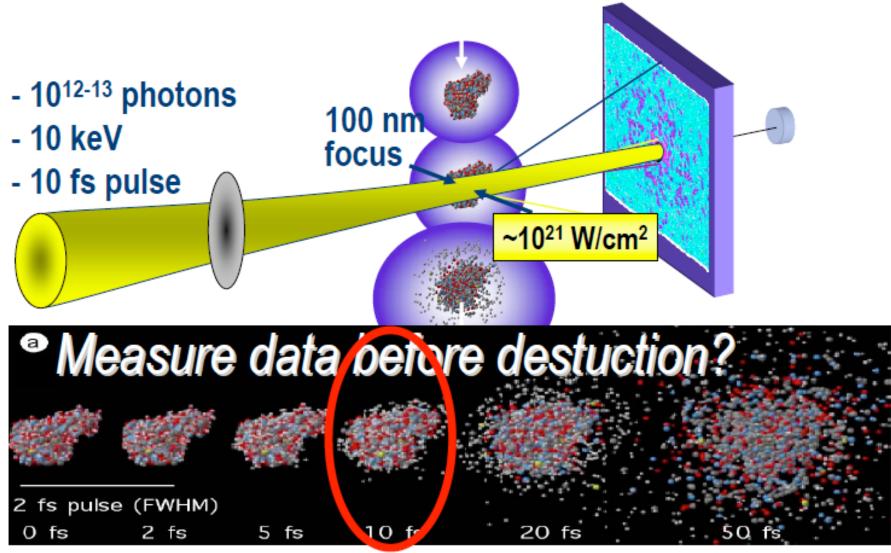
10¹² photons/pulse 900 J/cm²

Dose/pulse: 300-700 MGy

~ 7 µm diam. ~ 4 µm diam.

 $(z = 68 \text{ mm})_{1800 \text{ (3600) patterns/min}}$ 5 TB in one night

Coherent Diffractive Imaging



Calculations in vacuum, Neutze et al., Nature 2000

So what's the bad news?

- Hit rates are low, + only a fraction of hits indexable
- → the method needs:
 - 1-to-several ml of highly concentrated (yoghurt-like!) suspension of microcrystals (hit rates are low, for high resolution many 10,000s images needed)

```
How do you make that much protein?

(usual yields are in the 0.1-1mg range for membrane proteins..., very difficult to produce, not stable!)

Can you make nanocrystals of it?

(how do you know you have them?

how do you know they are any good?

testing them can only be done at the FEL...)

(can you inject them?

PEG/salts may clog the nozzle....)

DROPLET-ON-E
```

\$\$\$\$\$\$

DROPLET-ON-DEMAND TO SAVE SAMPLE ? HIGH PULSE RATE?

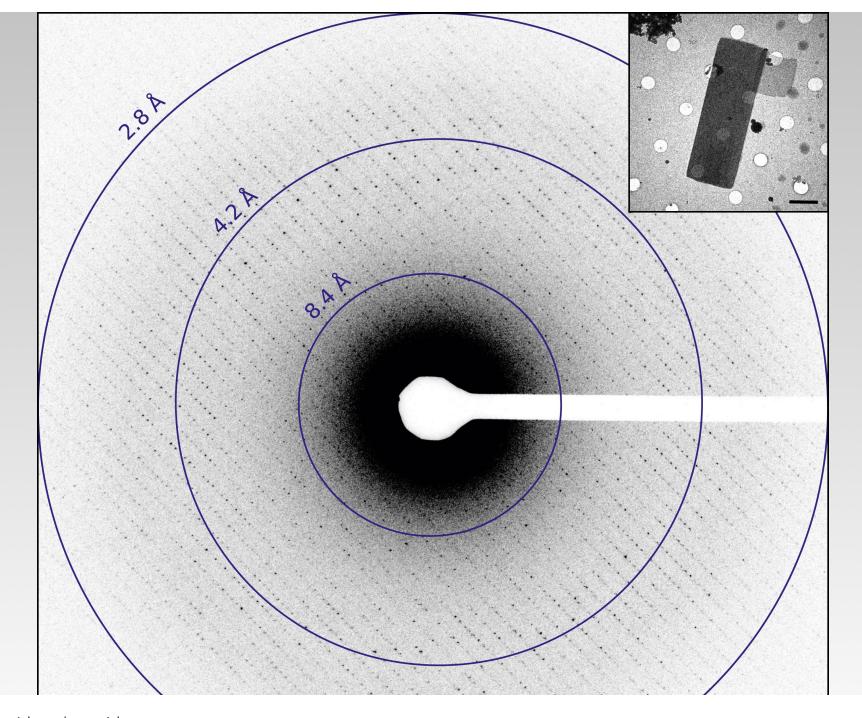
MicroED

• Microcrystal Electron Diffraction

uses a cryo-electron microscope--->

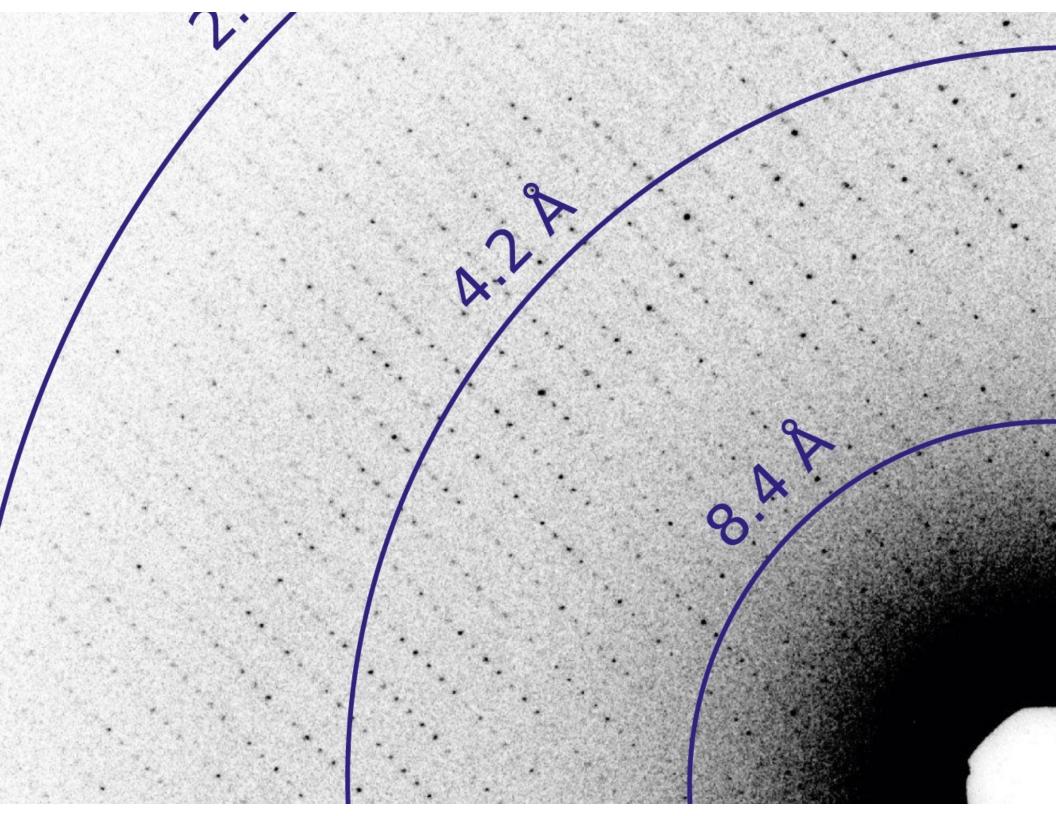
Proceed to Gonen2016.pdf



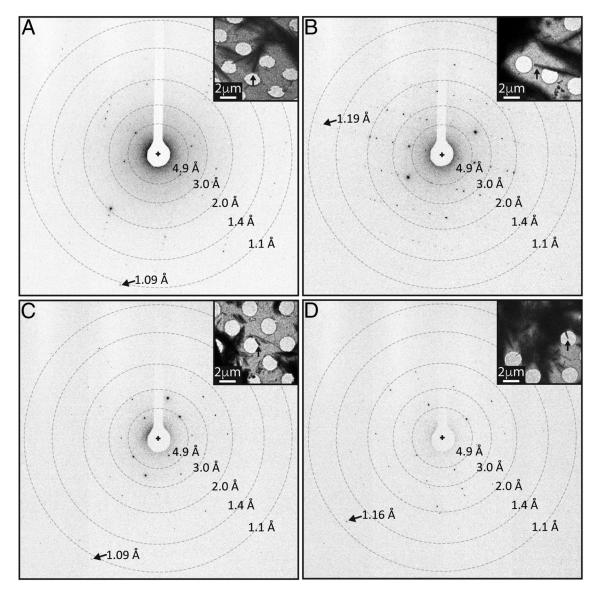


Diffraction from catalase microcrystals. Representative untilted still diffraction pattern of a catalase microcrystal that shows sharp reflections extending to approximately 3.0 Å. Crystals of this quality were used to collect a data set by continuous rotation. Inset shows an example catalase microcrystal as seen in over-focused diffraction mode. Scale bar is 2 μ m. The dimensions of most microcrystals varied between 6 and 20 μ m in length, 2 and 8 μ m in width, and the thickness was approximately 100–200 nm, in agreement with previous catalase crystal sizes (Dorset and Parsons, 1975b).

Figure 1.



Atomic resolution electron diffraction from amyloid nanocrystals.



Michael R. Sawaya et al. PNAS 2016;113:11232-11236

Comparison of latest crystallography methods

	XFEL	MicroED
crystals	micro/nano	nano
cost	\$\$\$\$	\$\$
protein	g	ng
scattering	e-	nucleus

Physics of Xray versus NMR

- In NMR <u>long</u>-wavelength light interacts with the <u>nuclei</u>.
 In Xray, <u>short</u> wavelength light interacts with the <u>electrons</u>.
- In NMR, light is an oscillating <u>magnetic</u> field.
 In Xray, light is an oscillating <u>electric</u> field.
- In NMR we Fourier transform from the <u>time</u> domain to the <u>frequency</u> domain (reciprocal time).
 In Xray we Fourier transform from the <u>space</u> domain to the <u>reciprocal space</u> domain.
- In NMR we solve a protein structure by determining the <u>internal</u> coordinates (pairwise distances).
 In Xray we solve a structure by determining the <u>Cartesian</u> coordinates.